Performance of GC-LRMS/MS and GCxGC methods for compliance monitoring of the PCDD/F-TEQ and the total TEQ in food and feed

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Introduction

Within the framework of the EU-DIFFERENCE project, an interlaboratory study was organized to assess the feasibility of certification of new reference materials for PCDD/F and dioxin-like PCB analysis in food and feed. In view of the Commission Directives 2002/69/EC and 2002/70/EC, consensus values and their uncertainties were based only on results obtained with the confirmatory method GC-HRMS. Because another objective of the DIFFERENCE project was the performance characterization of potentially cheaper and faster screening methods, some experienced GC-LRMS/MS and GCxGC laboratories were also invited to analyze the five test materials of the interlaboratory study. In this paper the bias and within-lab precision obtained with the various chemo-analytical methods is presented and discussed.

Materials and methods

Three food and two feed materials were selected and prepared for this study:

- a wet herring tissue, naturally contaminated (DIFF-01);

- a wet pork tissue, obtained by mixing regular meat with meat from a feeding experiment (DIFF-02);

- a whole milk, to which additional PCDD/Fs and dioxin-like PCBs were spiked (DIFF-03);

- a fish oil, naturally contaminated (DIFF-04);

- a compound feed for pigs, of which the lipid source was a salmon oil to which additional PCDD/Fs and dioxin-like PCBs were spiked (DIFF-05).

The materials were selected and prepared so that the levels in the final materials were close to the limits specified in the EC Directive 2001/102/EC and EC Regulation 2375/2001, except for the fish (with PCDD/F-TEQ level of about half the EC limit). The PCDD/F-TEQ and total TEQ, calculated from the GC-HRMS consensus values for the individual congeners after elimination of technically explainable outlying lab means, are given in Table 1. For the fish oil lower and upper bound TEQ were slightly different, so both are presented. The extractable fat content in Table 1 is the mean value obtained by the labs. Similarly, a moisture content in the compound feed of 9.3% was calculated. Table 1 : Material characteristics

Material	PCDD/F-TEQ	total TEQ	extractable fat
	(pg/g product)	(pg/g product)	(%)
Herring muscle tissue (DIFF-01)	1.89	3.76	17.6
Pork muscle tissue (DIFF-02)	0.307	0.446	23.6
Whole milk (DIFF-03)	0.0945	0.220	3.68
Fish oil (DIFF-04)	5.46-5.56	11.0-11.2	-
Compound feed (DIFF-05)	0.463	1.23	6.79

Details about the optimization of the screening methods as well as typical instrumental conditions have been published elsewhere¹⁻⁴. The GCxGC laboratories used ECD (lab 13) or TOFMS (lab 17) detection.

As guidelines for the evaluation, the requirements for analysis of PCDD/Fs and dioxin-like PCBs laid down in the Commission Directives 2002/69/EC and 2002/70/EC were applied, i.e.:

- trueness: -20 to +20% for confirmatory (GC-HRMS) methods

- CV_w: <30% for screening methods, <15% for confirmatory (GC-HRMS) methods.

The above criteria refer to the total TEQ value, but they were also used for the PCDD/F-TEQ value.

Results

The biases for the PCDD/F-TEQ and total TEQ observed with the chemical screening techniques are summarized in Figure 1.



Figure 1: Biases for PCDD/F-TEQ (upper) and total TEQ (lower) with GC-LRMS/MS and GCxGC methods.

It can be concluded that both GC-LRMS/MS and GCxGC (with ECD or TOFMS detection) can yield very good PCDD/F-TEQ and total TEQ estimates at the concentration levels investigated. With one exception all biases complied with the trueness criteria for a confirmatory method in the Commission Directives 2002/69/EC and 2002/70/EC, as did the biases of the GC-HRMS datasets used to calculate the congener consensus values. The GC-LRMS/MS data from lab 15 for fish and pork tissue were not retained because of an extraction problem identified at the stage of the technical evaluation.

Figure 2 shows the within-lab precision observed for the chemical screening techniques.



Figure 2: Within-lab precision for PCDD/F-TEQ (upper) and total TEQ (lower) with GC-LRMS/MS and GCxGC methods.

Only in one case the limit set for screening methods was slightly exceeded. The GC-LRMS/MS technique appeared able to achieve a similar precision as GC-HRMS, though the performance may strongly depend on the working conditions of the lab. On an overall basis, no clear relation between the within-lab precision and the matrix analyzed was found. It should be noted that the CV_w values obtained in this feasibility study most likely do not represent within-

lab reproducibility conditions completely.

Though the chemo-analytical methods investigated in this study can provide sufficiently accurate TEQ estimates, most of them may yield rather variable results for the mass fractions of individual congeners compared to the GC-HRMS confirmatory method, as illustrated in Figure 3. This is probably related to a higher limit of quantification and/or a more difficult peak integration. One of the GC-LRMS/MS labs (lab 15) reached a similar precision as the GC-HRMS labs in this feasibility study, but this required special precautions for instrument maintenance (e.g. frequent cleaning of the ion volume), which may not be sustainable in routine practice.

No Pooling - Lab Means & their C.I. for 1,2,3,4,7,8-H6CDF



Figure 3: Determination of 1,2,3,4,7,8-H₆CDF in whole milk DIFF-03;

L14 and L15 applied GC-LRMS/MS, L17 applied GCxGC-TOF

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