

1 Silicon accumulation in maize negatively impacts the feeding and life history traits of
2 *Spodoptera exigua* Hübner

3

4 Nicolas Leroy¹, Noé Hanciaux¹, Jean-Thomas Cornélis², and François J. Verheggen^{1,*}

5

6 ¹ Gembloux Agro-Bio Tech, TERRA, University of Liege, Avenue de la Faculté
7 d'Agronomie 2B, 5030 Gembloux, Belgium

8 ² Water-Soil-Plant Exchanges, Gembloux Agro-Bio Tech, University of Liege, Avenue
9 Maréchal Juin 27, 5030 Gembloux, Belgium

10 * Corresponding authors: fverheggen@uliege.be

11 Abstract:

12 Silicon (Si) accumulation in plant tissues helps alleviate abiotic and biotic stresses, including
13 infestation by insect pests. Here, we tested the hypothesis that Si concentration in maize leaves
14 negatively impacts *Spodoptera exigua* Hübner (Lepidoptera: Noctuidae) with respect to: (i)
15 feeding preferences; (ii) leaf digestion; and (iii) life history traits. We produced three groups of
16 maize plants cultivated in a hydroponic system that had contrasting Si concentrations in their
17 tissues (i.e., 0.21 ± 0.03 , 4.45 ± 0.50 and 8.46 ± 0.61 g Si Kg⁻¹ DW). In choice assays, fifth
18 instars preferentially consumed leaves containing lower Si concentrations. In no choice-assays,
19 we found that the approximate digestibility (AD) of larvae feeding on Si-enriched leaves was
20 not affected. However, these larvae exhibited a 32% reduction in relative growth rates. Higher
21 Si concentration in maize leaves extended larval development by three days; from 18.07 ± 0.29
22 when feeding on Si- diet to 21.39 ± 0.21 days on the Si⁺⁺ enriched diet. Silicon also reduced
23 larval survival by 18% and pupal weight by 20%. Our results confirm that Si supplementation
24 in soil enhances the ability of plants to resist infestation with chewing insects, and should be
25 considered as a viable option in the existing range of sustainable management practices.

26

27 Keywords: beet armyworm, *Zea mays*, silicon, herbivory, plant resistance, feeding preference,
28 Noctuidae

29

30 **Introduction**

31 Silicon (Si) is among the most abundant elements in terrestrial environments. Plant tissues are
32 no exception, as they contain up to 10% Si, depending on species accumulation ability (Epstein,
33 1994; Richmond & Sussman, 2003; Hodson *et al.*, 2005). While not considered essential for
34 plant development, Si has been extensively studied for its action on plant growth, yield and
35 quality (Etesami & Jeong, 2018; Guo-chao *et al.*, 2018). In addition, recent reports document
36 the importance of Si uptake and accumulation on the ability of plants to alleviate abiotic and
37 biotic stresses, including infestation by pests and infection by pathogens (Ma, 2004; Liang *et*
38 *al.*, 2015; Reynolds *et al.*, 2016; Alhousari & Greger, 2018; de Tombeur *et al.*, 2020).

39 More precisely, Si increases the resistance of plants against insect herbivory via several
40 mechanisms (Kvedaras *et al.*, 2007; Reynolds *et al.*, 2009). For instance, Si accumulates as
41 amorphous hydrated silica ($\text{SiO}_2 \cdot n\text{H}_2\text{O}$), increasing the abrasiveness and hardness of plant
42 tissue. Subsequently, it damages the mandibles and other oral parts of insects, as well as
43 reducing the digestibility, palatability, and overall consumption of plants (Jeer *et al.*, 2017).
44 Higher Si concentrations allow plants to develop silica-rich defensive structures, including
45 trichomes or prickle cells (Andama *et al.*, 2020; Hall *et al.*, 2020). Si also impacts the nutritional
46 quality of plants (Frew *et al.*, 2019). The biochemical responses of plants against insect
47 infestation might also be promoted by Si (Rahman *et al.*, 2015). Plants characterized by high Si
48 accumulation exhibit higher expressions of genes encoding for defensive enzymes, resulting in
49 altered levels of defensive compounds (Gomes *et al.*, 2005; Rémus-Borel *et al.*, 2005; Yang *et*
50 *al.*, 2017).

51 While integrated pest management strategies are emerging as a response to stricter regulations
52 on insecticides, Si supplementation represents a possible alternative measure to limit damage
53 from insect pests in several agricultural crops (Bakhat *et al.*, 2018; Leroy *et al.*, 2019; de
54 Tombeur *et al.*, 2021). Evidence is accumulating on the beneficial role of Si supplements in

55 various plant-pest associations. For instance, significant damage reduction by a stemborer
56 (*Chilo suppressalis* Walker) and a leaf folder (*Cnaphalocris medinalis* Guenée) was observed in
57 fields containing rice under Si amendment (Hou & Han, 2010; Han *et al.*, 2015). In field-grown
58 sugarcane, Si treatment significantly reduced the proportion of stalks bored by *Eldana*
59 *saccharina* Walker (Keeping *et al.*, 2013). The use of Si in controlling *Spodoptera* sp.
60 populations has also received noticeable research interest. However, studies have mainly
61 focused on *S. frugiperda* and *S. exempta* (to a lesser extent). Si accumulation in maize and
62 cotton plants adversely affects the survival of *S. frugiperda* larvae (Goussain *et al.* 2002; Silva
63 *et al.*, 2014). Si also affects the life history traits of *S. frugiperda*, including larval weight, adult
64 longevity, and fertility (Nascimento *et al.*, 2014; Alvarenga *et al.*, 2017).

65 Here, we tested the hypothesis that Si concentration in maize leaves negatively impacts
66 *Spodoptera exigua* Hübner with respect to: (i) feeding preferences; (ii) leaf digestion; and (iii)
67 life history traits. We cultivated maize plants in a hydroponic system to produce three groups
68 with different Si concentrations. The impact of Si in plant tissue on the ability of a plant to
69 alleviate *S. exigua* Hübner (Lepidoptera: Noctuidae) damage has not been previously studied,
70 despite its worldwide economic pest status (Zheng *et al.*, 2011). Based on studies performed on
71 other armyworms (i.e. *S. frugiperda* and *S. exempta*) (Massey & Hartley, 2006; Alvarenga *et*
72 *al.*, 2017), we hypothesized that higher Si assimilation in maize tissue would reduce the survival
73 of pupae and larvae. We also expected other life history traits (including larval weight and pupal
74 weight) to be reduced on plants with higher concentrations of Si. By demonstrating the positive
75 impact of Si supplementation on the ability of maize plants to combat *S. exigua* infestations
76 would expand the range of available options for sustainable control management.

77 **Material and methods**

78 *Plant material and hydroponic system* – Corn plants (*Zea mays* L. var Delprim) were used in
79 all experiments (Delley Seeds and Plants, Delley, Switzerland). Seeds were germinated on
80 paper towels moistened with distilled water in Petri dishes for three days under dark conditions
81 at 23 °C. Seedlings were transplanted to rockwool substrate, inside a 20-litre plastic bucket
82 containing water, and placed in a temperate chamber set at: 23 ± 2 °C (day), 19 ± 2 °C (night),
83 55-70% relative humidity (RH), and 300 µmol.m⁻².s⁻¹ light intensity. After four days, water was
84 replaced by a commercial nutrient solution (HY-PRO[®] A&B, Bladel, Netherlands) [46.29 mg.l⁻¹
85 ¹ N; 23,94 mg.l⁻¹ P; 227.81 mg.l⁻¹ K; 115.12 mg.l⁻¹ Ca; 0.09 mg.l⁻¹ Cu; 38.79 mg.l⁻¹ Mg; 1.48
86 mg.l⁻¹ Fe; 0.15 mg.l⁻¹ Mn; 0.13 mg.l⁻¹ Zn; 3.71 mg.l⁻¹ Na]. Continuous aeration was maintained
87 in the plastic bucket using an air pump. The seedlings were grown in quarter strength nutrient
88 (diluted 4x) solution for two days, with the solution being gradually raised to full strength over
89 one week to avoid osmotic shock. The nutrient solution was renewed every three days. The pH
90 of the medium was adjusted to 5.5 ± 0.5 by adding 0.5 M MgSO₄. This addition also corrected
91 the Mg/K ratio in the nutrient solution and prevented Mg deficiency in maize plants. One week
92 after being cultivated in this nutrient solution, all plants were separated to one of the three Si
93 concentrations: (a) control solution without Si addition, named Si⁻ [0.05 mM Si]; (b) medium
94 level of Si, named Si⁺ [0.6 mM Si]; (c) a highly-enriched solution, named Si⁺⁺ [2.0 mM Si].
95 Si was supplied as monosilicic acid (H₄SiO₄) in the nutrient solution. The concentration of the
96 Si⁺ solution was chosen according to the average concentration of Si found in soil (Epstein,
97 1994). The concentration of the Si⁺⁺ solution was set according to the limit of saturation of
98 silicic acid (> 2mM), at which point it precipitates as amorphous silica (Exley, 2015). The
99 monosilicic acid solution was freshly prepared by dissolving sodium metasilicate in
100 demineralized water, and passing the solution through cation-exchange resin (Amberlite[®] IR-
101 120) (Cornelis *et al.*, 2010).

102 *Insect rearing* – Beet armyworm *S. exigua* eggs were purchased from Entocare Biological
103 Control (Wageningen, Netherlands). After three days of incubation at 24 °C, first instars were
104 transferred to an artificial diet (General purpose Lepidoptera, Frontier Scientific Services
105 Agriculture, Newark, USA). The insects were reared at 24 ± 2 °C and 40–50% relative
106 humidity, under a 18:6 (L:D) photoperiod.

107 *Foliar Si content* – Foliar Si content was determined on maize plants that were grown for 30 to
108 35 days in the hydroponic system (17-18 BBCH growth stage). All leaves were collected from
109 one plant, dried at 50 °C for 72 h, ground in a plant shredder, and left for 24 h at 450 °C for
110 calcination. One hundred milligrams of ash was melted at 1000 °C for 5 min in a graphite
111 crucible containing 0.4 g Li-tetraborate and 1.6 g Li-metaborate (Chao & Sanzalone, 1992).
112 The fusion bead was then dissolved in 10% HNO₃ before quantifying Si concentrations by ICP-
113 OES. Five quantifications were performed per Si treatment, using different plants.

114 *Effect of Si accumulation on the feeding preference of S. exigua* – We aimed to evaluate the
115 feeding preferences of *S. exigua* larvae in the presence of a piece of three maize leaves of
116 contrasting Si concentrations. One entire maize leaf (5th and 6th leaf) was collected from each
117 group of plants (Si-, Si+, Si++) using a pair of scissors. All three leaves were washed and cut
118 in 5x3 cm pieces. One piece of each leaf was placed equidistantly in a glass Petri dish (20 cm
119 diameter) containing a 1% agar-water layer. One caterpillar (third or fifth instar) was deposited
120 in the centre of the Petri dish and the arena was sealed with PVC film. The assay was repeated
121 54 and 50 times, for third and fifth instars, respectively. All Petri dishes were placed in a dark
122 room to prevent the effect of phototropism. Leaf area was recorded before caterpillar infestation
123 after 48 h using ImageJ Software (Rasband 1997-2015). The consumed leaf area was calculated
124 by subtraction (Nascimento et al. 2017). For most replicates, in each treatment, the piece of leaf
125 was not entirely consumed. The replicate was removed if such case arose. The bioassay was
126 conducted at 23 °C and 50% RH.

127 *Effect of Si accumulation on the consumption and digestion of leaves by S. exigua* – Fifth instars
128 were removed from the artificial diet and starved for 24 h. They were then weighed before being
129 placed in separate Petri dishes (9 cm diameter) and offered a piece of maize leaf (5x3 cm), like
130 those used in the previous experiment. Larvae were allowed to feed for 72 h, and were then
131 starved again for 24 h to allow the faeces to be evacuated, before being weighed again. All frass
132 (excrement) was collected, dried and weighed. Leaves were renewed every day, and the
133 remaining leaf material was dried at 50 °C and weighed. Relative growth rate (RGR) was
134 calculated as mass gained (mg)/initial mass (mg) * time (days), over the three days of the
135 experiment (Hall et al. 2020). We also evaluated Approximate digestibility (AD) as the weight
136 of food ingested (mg dry mass) – weight of faeces (mg dry mass) / weight of food ingested (mg
137 dry mass) * 100 (Massey & Hartley, 2009).

138 *Effect of Si accumulation on the life history traits of S. exigua* – Neonates were placed in
139 separate Petri dishes (9 cm diameter) filled with a 1 % agar-water layer, and were fed *ad libitum*
140 with 5x3 cm pieces of maize leaves cut from Si-, Si+ and Si++ plants. Insects were housed at
141 23 °C and 50 % RH. Leaves were changed every two days. Various life history traits were
142 collected until adult emergence, including larval mortality, stage duration, larval size, larval
143 weight, pupal mortality, pupal stage duration, pupal weight and sex-ratio of the pupae.

144 *Statistical analyses* – R studio software (v 1.2.1335) was used for all statistical analyses (R core
145 team 2019). Data on Si content had to be square root transformed. Data on feeding preference
146 with third and fifth instars, larval stage duration and pupal weight were transformed using
147 (rn)transform function (GenAbel package). Data on RGR and the growth in size of larvae were
148 not transformed. Data were transformed to obtain a normal distribution. Statistical analyses
149 associated with the growth in size of larvae was carried out by comparing the slope of each line
150 associated with the growth of each individual over time. Data were subjected to analysis of
151 variance (ANOVA) and Tukey's post-hoc test ($\alpha = 0.05$). Data on larval survival, pupal survival

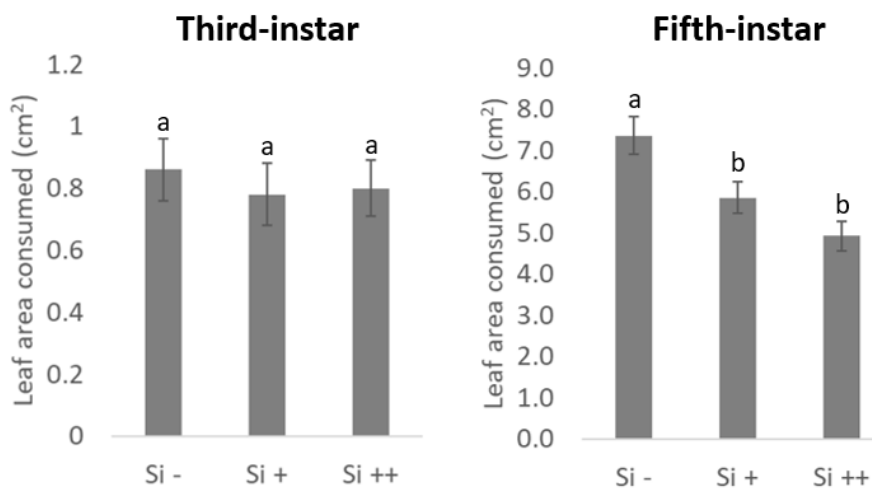
152 and sex-ratio were subjected to generalised linear models (GLM), assuming a binomial
153 distribution. We could not transform the data on the duration of the pupal stage and approximate
154 digestibility (AD) to meet the requirements of the analysis of variance. Therefore, a Kruskal-
155 Wallis test was used on these parameters.

156

157 **Results**

158 *Foliar Si content* – The Si content of leaves was dependent on the Si concentration of the
159 nutrient solution ($F_{2,12} = 190.8$; $P < 0.001$). We recorded the following mean silicon
160 concentrations: 0.213 ± 0.030 g Kg⁻¹DW (Si-), 4.451 ± 0.498 g Kg⁻¹DW (Si+) and $8.459 \pm$
161 0.611 g Kg⁻¹DW (Si++) (mean \pm SD). The means of all three concentrations significantly
162 differing from each other (Tukey Post-Hoc test, $P < 0.05$).

163 *Effect of Si accumulation on the feeding preferences of S. exigua* – In choice assays, fifth instars
164 preferentially consumed leaves with lower Si concentrations ($F_{2,159} = 11.23$; $P < 0.001$) (Fig. 1
165 B). Instars consumed 33% more Si- leaf material compared to Si++ material. This difference
166 was not observed for third instars ($F_{2,155} = 0.174$; $P = 0.841$) (Fig. 1 A).

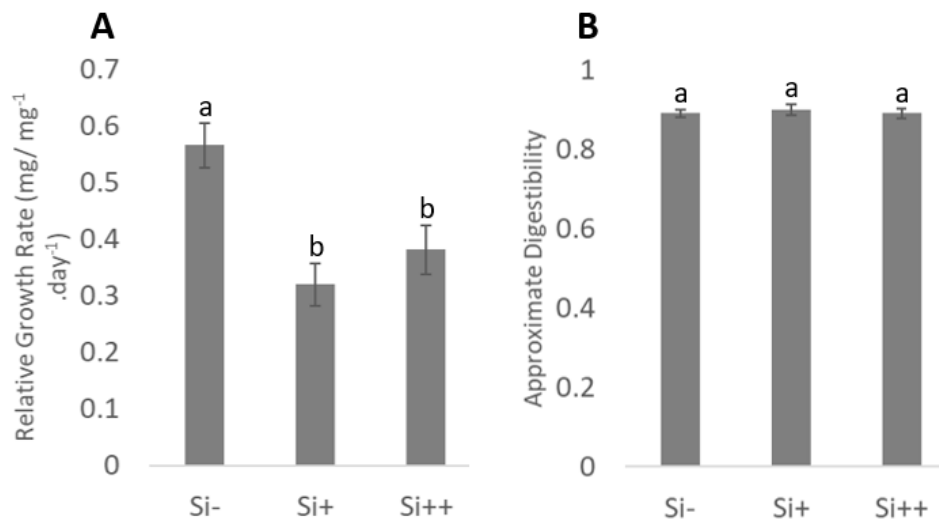


167

168 Fig. 1. Leaf area consumed by *S. exigua* third instar larvae (n=54) and fifth instar larvae (n =
169 50) in choice assays. Values are means \pm SD. Different letters on bars indicate significant
170 differences by Tukey's test ($p \leq 0.05$).

171

172 *Effect of Si accumulation on the consumption and digestion of leaves by S. exigua* – In the no-
173 choice assays, the relative growth rate of instars was negatively impacted ($F_{2,89} = 10.14$; $P <$
174 0.001) (Fig. 2) Concerning food-utilization measurement, we found no significant difference
175 between each Si treatment ($\chi^2 = 0.671$; $df = 2$, $P = 0.71$).



176

177 Fig. 2. Relative growth rate (A), Approximate digestibility (B) by *S. exigua* fifth instars (n =
178 30 per treatments). Values are means \pm SD. Different letters on bars indicate no difference by
179 Tukey's test ($p < 0.05$).

180 *Effect of Si accumulation on the life history traits of S. exigua* – Si concentration in maize leaves
181 impacted some of the recorded life history traits of *S. exigua*. We found that elevated Si
182 concentrations extended the duration of larval development ($F_{2,65} = 34.84$; $P < 0.001$), leading
183 to lighter pupae ($F_{2,65} = 8.51$; $P < 0.001$) (Tab. 1). While 58% of larvae survived under Si-, just
184 42% and 40% became pupae under Si+ and Si++ concentrations, respectively ($F_{2,147} = 2.62$; P

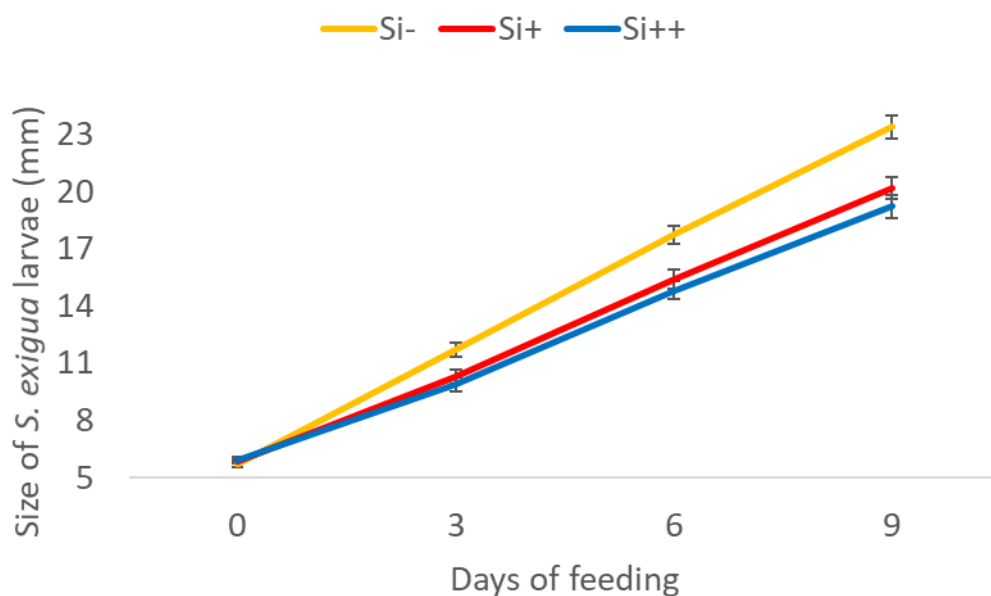
185 = 0.073). Si concentration in maize leaves significantly impacted the development of *S. exigua*
 186 larvae ($F_{2,74} = 12.74$; $P < 0.001$) (Fig. 3). Differences were highlighted between Si- and Si+
 187 treatments ($P < 0.0001$) and between Si- and Si++ treatments ($P < 0.0001$). The development
 188 of larvae exposed to Si+ and Si++ was not significantly different ($P=0.743$). We found that Si
 189 concentration did not impact the duration of the pupal stage ($\chi^2 = 0.216$; $df = 2$, $P = 0.89$), nor
 190 the survival of pupae ($F_2 = 0.093$; $P = 0.911$), nor sex-ratio ($F_2 = 0.415$; $P = 0.661$).

191 Table 1. Impact of Si concentration in maize leaves on *S. exigua* life history traits. Different
 192 letters on means and percentages indicate significant differences

Si treatments	Larval stage duration (days)	Larval survival (%)	Pupal weight (g)	Pupal stage duration (days)	Pupal survival (%)	Sex ratio (M/F)
Si-	18.07 ± 0.29 ^a	58 ^a	75.54 ± 1.62 ^a	9.41 ± 0.14 ^a	76 ^a	0.94 ^a
Si+	20.67 ± 0.27 ^b	42 ^a	68.01 ± 3.35 ^{ab}	9.42 ± 0.21 ^a	81 ^a	0.75 ^a
Si++	21.39 ± 0.21 ^b	40 ^a	60.89 ± 2.69 ^b	9.57 ± 0.27 ^a	78 ^a	0.64 ^a

193

194



195

196 Fig. 3. *S. exigua* larvae size growth over the third instar (day 0 corresponds to the start of the
197 third instar)

198

199 **Discussion**

200 Previous studies have shown that Si enhances the ability of plants to resist insect pests (Luyckx
201 *et al.* 2017; Islam *et al.*, 2020; Johnson *et al.*, 2020). In this study, we demonstrated that Si
202 accumulation in maize plants negatively impacts some life history traits of *S. exigua*. High Si
203 concentrations extended the duration of the larval stage and reduced pupal weight. We also
204 found an 18% increase in larval mortality; however, this result was not supported statistically.

205 Pupal weight declined following exposure to plants containing Si, supporting a previous study
206 on *S. exempta* (Massey & Hartley 2006). However, contrasting results were obtained for *S.*
207 *frugiperda*. Some studies recorded mortality in *S. frugiperda* caterpillars fed with silicon-
208 enriched diets, while other studies recorded no impact on life history traits (Silva *et al.*, 2014;
209 Alvarenga *et al.*, 2017). Nascimento *et al.* (2017) found that Si addition to rice plants only
210 reduced the weight of larvae and pupae. Nagaratna *et al.* (2021) observed that biological
211 parameters of *S. frugiperda* such as larval weight and larval survival were negatively impacted
212 by Si application. Si deposition as phytoliths within plant tissue is likely one of the main factors
213 explaining the increased mortality of caterpillars. Phytoliths reduce the digestibility and
214 palatability of plants by increasing rigidity and abrasiveness (Strömberg *et al.*, 2016). The other
215 possible mechanism is linked to the biochemical response of plants against pests. There is
216 increasing evidence that Si treatment alters the accumulation of various defensive compounds,
217 including phytoalexins, phenolics and chlorogenic acid (Rémus-Borel *et al.*, 2005; Rahman *et*
218 *al.*, 2015; Frew *et al.*, 2016; Wang *et al.*, 2021; de Tombeur *et al.*, 2021).

219 Unlike third instars, fifth instars preferentially consume maize leaves with low Si
220 concentrations. Insects consuming phytolith-rich diets experience wear on the mandibles
221 (Massey & Hartley, 2009), which could explain the selection of non-enriched leaves. Again,
222 this conclusion is only partially supported by the published literature on *Spodoptera* (Massey
223 *et al.*, 2006; Nascimento *et al.*, 2014). In our study, we regularly checked the content of the
224 Petri dish, and we observed that all fifth instars tested all three pieces of leaves before
225 preferentially feeding on the less Si-concentrated one. Our data suggest that Si acts as a feeding
226 deterrent, and that fifth larvae are able to perceive this deterrent effect when feeding. However,
227 this effect was not observed in third instars. Given their smaller size, it is possible that the
228 quantity of leaf consumed was not sufficient to allow us to compare diets (third instars
229 consumed 10 times less leaf mass than fifth instars).

230 Silica negatively impacts the digestibility of leaves by *S. exempta*, as demonstrated previously
231 on various grass species (*Deschampsia caespitosa* L.; *Festuca ovina* L. and *Lolium perenne* L.)
232 by Massey & Hartley (2009). The authors concluded that silica has a progressive impact with
233 longer exposure time, indicating that herbivores cannot adapt to silica defences, and that they
234 do not develop tolerance for silica with age. We also observed a reduction in the relative growth
235 rate of *S. exigua* that fed, even for a short time, on high Si-enriched maize leaves. Whether this
236 effect increases with exposure duration remains to be tested for *S. exigua*.

237 The effect of Si-accumulation in plant leaves on the life history traits of *Spodoptera* sp. in our
238 study supported some existing studies and contrasted with others. Supporting previous studies,
239 we found that several traits were not affected, including pupal stage duration, pupal survival,
240 sex-ratio and approximate digestibility (AD). Thus, Si-stimulated defence mechanisms are not
241 as efficient against *Spodoptera* spp. as other lepidopterans (Keeping & Meyer, 2013; Johnson
242 *et al.*, 2020). To describe the underlying mechanisms, future studies should evaluate the levels
243 of defensive compounds, focusing on the interplay between jasmonic acid (JA) (a

244 phytohormone that regulates chemical defences against chewing insects) and Si accumulation
245 (Johnson *et al.*, 2021; Waterman *et al.*, 2021). Specifically, DIMBOA (2,4-dihydroxy-7-
246 methoxy-1,4-benzoxazin-3-one) has antifeedant effect on *S. exigua* (Rostás, 2007). As the
247 feeding preferences of *S. exigua* are impacted by Si accumulation in maize leaves, future studies
248 should investigate the impact of Si concentrations in leaves on the level of DIMBOA content.

249 Si concentrations of up to 0.85% were reached in our assays. These concentrations were similar
250 to those used in previous studies (e.g. Johnson *et al.*, 2020), which also reported Si-stimulated
251 herbivore resistance, except for in non-Poaceae species. The Si concentrations reached in the
252 maize leaves in the current study were comparatively low to that found in other varieties of
253 maize, with ranges of 1% to 2.5% (Ma *et al.*, 2001; Liang *et al.*, 2015). Thus, even higher Si
254 concentrations could be added to maize, which would potentially enhance our results.

255 Our results confirmed that Si supplementation to the soil increases absorption by plants and the
256 subsequent ability of plants to resist infestation with chewing insects. This approach should be
257 considered among the diversity of available options of sustainable management practices for
258 crop plants. However, future studies must focus on elucidating the mechanisms of action of Si
259 on chewing insects. Also, field assays should be formulated to evaluate the beneficial role of Si
260 through soil fertilization or by optimizing the Si biological cycle under outdoor conditions.

261

262 **Acknowledgments**

263 Leroy Nicolas is supported by a FNRS-FRIA grant.

264

265 **References**

- 266 Alhousari, F., & Greger, M. (2018). Silicon and mechanisms of plant resistance to insect pests.
267 *Plants* 7(2), 33. <https://doi.org/10.3390/plants7020033>
- 268 Alvarenga, R., Moraes, J.C., Auad, A.M., Coelho, M., & Nascimento, A.M. (2017). Induction
269 of resistance of corn plants to *Spodoptera frugiperda* (J.E. Smith, 1797) (Lepidoptera :
270 Noctuidae) by application of silicon and gibberellic acid. *Bulletin of Entomological Research*
271 107, 527–533. <https://doi.org/10.1017/S0007485316001176>
- 272 Andama, J.B., Mujiono, K., Hojo, Y., Shinya, T., & Galis, I. (2020). Nonglandular silicified
273 trichomes are essential for rice defense against chewing herbivores. *Plant Cell and Environment*
274 104, 2019–2032. <https://doi.org/10.1111/pce.13775>
- 275 Bakhat, H.F., Bibi, N., Zia, Z., Abbas, S., Hammad, H.M., Fahad, S., Ashraf, M.R., Shah, G.M.,
276 Rabbani, F., & Saeed, S. (2018). Silicon mitigates biotic stresses in crop plants: A review. *Crop*
277 *Protection* 104, 21–34. <https://doi.org/10.1016/j.cropro.2017.10.008>
- 278 Cornelis, J.T., Delvaux, B., & Titeux, H. (2010). Contrasting silicon uptakes by coniferous
279 trees: A hydroponic experiment on young seedlings. *Plant and Soil* 336, 99–106.
280 <https://doi.org/10.1007/s11104-010-0451-x>
- 281 de Tombeur, F., Vander Linden, C., Cornélis, J.T., Godin, B., Compère, P., & Delvaux, B.
282 (2020). Soil and climate affect foliar silicification patterns and silica-cellulose balance in
283 sugarcane (*Saccharum officinarum*). *Plant and Soil* 452, 529–546.
284 <https://doi.org/10.1007/s11104-020-04588-z>
- 285 de Tombeur, F., Cooke, J., Collard, L., Cisse, D., Saba, F., Lefebvre, D., Burgeon, V., Nacro,
286 H.B., & Cornélis, J.T. (2021). Biochar affects silicification patterns and physical traits of rice
287 leaves cultivated in a desilicated soil (Ferric Lixisol). *Plant and Soil* 460, 375-390.
288 <https://doi.org/10.1007/s11104-020-04816-6>
- 289 de Tombeur, F., Laliberté, E., Lambers, H., Faucon, M.P., Zemunik, G., Turner, B.L., Cornélis,
290 J.T., & Mahy, G. (2021). A shift from phenol to silica-based leaf defenses during long-term soil
291 and ecosystem development. *Ecology letters* 24(5),984-995. <https://doi.org/10.1111/ele.13713>
- 292 Epstein, E. (1994). The anomaly of silicon in plant biology. *Proceedings of the National*
293 *Academy of Sciences* 91, 11–17. <https://doi.org/10.1073/pnas.91.1.11>
- 294 Etesami, H., & Jeong, B.R. (2018). Silicon (Si): Review and future prospects on the action
295 mechanisms in alleviating biotic and abiotic stresses in plants. *Ecotoxicology and*
296 *Environmental Safety* 147, 881–896. <https://doi.org/10.1016/j.ecoenv.2017.09.063>
- 297 Exley, C. (2015). A possible mechanism of biological silicification in plants. *Frontiers in Plant*
298 *Science* 6,853. <https://doi.org/10.3389/fpls.2015.00853>
- 299 Frew, A., Powell, J.R., Sallam, N., Allsopp, P.G., & Johnson, S.N. (2016). Trade-offs between
300 silicon and phenolic defenses may explain enhanced performance of root herbivores on
301 phenolic-rich plants. *Journal of chemical ecology* 42, 768-771. [https://doi.org/10.1007/s10886-](https://doi.org/10.1007/s10886-016-0734-7)
302 [016-0734-7](https://doi.org/10.1007/s10886-016-0734-7)
- 303 Frew, A., Weston, L.A., & Gurr, G.M. (2019). Silicon reduces herbivore performance via
304 different mechanisms, depending on host–plant species. *Austral Ecology* 44, 1092– 1097.
305 <https://doi.org/10.1111/aec.12767>

306 Gomes, F.B., de Moraes, J.C., dos Santos, C.D., & Goussain, M.M. (2005). Resistance
307 induction in wheat plants by silicon and aphids. *Scientia Agricola* 62, 547–551.

308 Goussain, M.M., Moraes, J.C., Carvalho, J.G., Nogueira, N.L., & Rossi, M.L. (2002). Effect of
309 silicon application on corn plants upon the biological development of the fall armyworm
310 *Spodoptera frugiperda* (J.E. Smith) (Lepidoptera: Noctuidae). *Neotropical entomology* 31(2),
311 305-310.

312 Guo-chao, Y., Nikolic, M., Mu-jun, Y., Zhuo-xi, X., & Yong-chao, L. (2018). Silicon
313 acquisition and accumulation in plant and its significance for agriculture. *Journal of Integrative*
314 *Agriculture* 17, 2138–2150. [https://doi.org/10.1016/S2095-3119\(18\)62037-4](https://doi.org/10.1016/S2095-3119(18)62037-4)

315 Hall, C.R., Dagg, V., Waterman, J.M., & Johnson, S.N. (2020). Silicon alters leaf morphology
316 and suppresses insect herbivory in a model grass species. *Plants* 9(5), 643.
317 <https://doi.org/10.3390/plants9050643>

318 Han, Y., Lei, W., Wen, L., & Hou, M. (2015). Silicon-mediated resistance in a susceptible rice
319 variety to the rice leaf folder *Cnaphalocris medinalis* Guenée (Lepidoptera: Pyralidae). *PLoS*
320 *ONE* 10(4), e0120557. <https://doi.org/10.1371/journal.pone.0120557>

321 Hodson, M.J., White, P.J., Mead, A., & Broadley, M.R. (2005). Phylogenetic variation in the
322 silicon composition of plants. *Annals of Botany* 96, 1027–1046.
323 <https://doi.org/10.1093/aob/mci255>

324 Hou, M., & Han, Y. (2010). Silicon-mediated rice plant resistance to the Asiatic rice borer
325 (Lepidoptera: Crambidae): Effects of silicon amendment and rice varietal resistance. *Journal*
326 *of Economic Entomology* 103(4), 1412-1419. <https://doi.org/10.1603/EC09341>

327 Islam, W., Tayyab, M., Khalil, F., Hua, Z., Huang, Z., & Chen, H.Y.H. (2020). Silicon-
328 mediated plant defense against pathogens and insect pests. *Pesticide Biochemistry and*
329 *Physiology* 168, 104641. <https://doi.org/10.1016/j.pestbp.2020.104641>

330 Jeer, M., Telugu, U.M., Voleti, S.R., & Padmakumari, A.P. (2017). Soil application of silicon
331 reduces yellow stem borer, *Scirpophaga incertulas* (Walker) damage in rice. *Journal of Applied*
332 *Entomology* 141, 189–201. <https://doi.org/10.1111/jen.12324>

333 Johnson, S.N., Rowe, R.C., & Hall, C.R. (2020). Silicon is an inducible and effective herbivore
334 defence against *Helicoverpa punctigera* (Lepidoptera: Noctuidae) in soybean. *Bulletin of*
335 *Entomological Research* 110, 417-422. <https://doi.org/10.1017/S0007485319000798>

336 Johnson, S.N., Hartley, S.E., Ryalls, J.M.W., Frew, A., & Hall, C.R. (2021). Targeted plant
337 defense: silicon conserves hormonal defense signaling impacting chewing but not fluid-feeding
338 herbivores. *Ecology* 102(3), e03250. <https://doi.org/10.1002/ecy.3250>

339 Keeping, M.G., Meyer, J.H., & Sewpersad, C. (2013). Soil silicon amendments increase
340 resistance of sugarcane to stalk borer *Eldana saccharina* Walker (Lepidoptera: Pyralidae) under
341 field conditions. *Plant and Soil* 363, 297–318. <https://doi.org/10.1007/s11104-012-1325-1>

342 Kvedaras, O.L., Keeping, M.G., Goebel, F.R., & Byrne, M.J. (2007). Larval performance of
343 the pyralid borer *Eldana saccharina* Walker and stalk damage in sugarcane: Influence of plant
344 silicon, cultivar and feeding site. *International Journal of Pest Management* 53, 183–194.
345 <https://doi.org/10.1080/09670870601110956>

346 Leroy, N., de Tombeur, F., Walgraffe, Y., Cornélis, J.-T., & Verheggen, F.J. (2019). Silicon and
347 plant natural defenses against insect pests : impact on plant volatile organic compounds and
348 cascade effects on multitrophic interactions. *Plants* 8(11),444.

349 <https://doi.org/10.3390/plants8110444>

350 Luyckx, M., Hausman, J.-F., Lutts, S., & Guerriero, G. (2017). Silicon and plants: Current
351 knowledge and technological perspectives. *Frontiers in Plant Science* 8, 411.
352 <https://doi.org/10.3389/fpls.2017.00411>

353 Liang, Y., Nikolic, M., Bélanger, R.R., Gong, H., & Song, A. (2015). *Silicon in Agriculture.*
354 *From theory to practice.* Berlin, Germany.

355 Ma, J.F., Miyake, Y., & Takahashi, E. (2001). Silicon as a beneficial element for crop plants.
356 In Datnoff, L.E., Snyder, G.H., & Korndöfer, G.H. (ed.), *Silicon in Agriculture*, Elsevier
357 Science, pp 17-39.

358 Ma, J.F. (2004). Role of silicon in enhancing the resistance of plants to biotic and abiotic
359 stresses. *Soil Science and Plant Nutrition* 50, 11–18.
360 <https://doi.org/10.1080/00380768.2004.10408447>

361 Massey, F.P., Ennos, A.R., & Hartley, S.E. (2006). Silica in grasses as a defence against insect
362 herbivores: Contrasting effects on folivores and a phloem feeder. *Journal of Animal Ecology*
363 75, 595–603. <https://doi.org/10.1111/j.1365-2656.2006.01082.x>

364 Massey, F.P., & Hartley, S.E. (2009). Physical defences wear you down : progressive and
365 irreversible impacts of silica on insect herbivores. *Journal of Animal Ecology* 78, 281–291.
366 <https://doi.org/10.1111/j.1365-2656.2008.01472.x>

367 Nagaratna, W., Kalleshwaraswamy, C.M., Dhananjaya, B.C., Sharanabasappa, & Prakash, N.B.
368 (2021). Effect of silicon and plant growth regulators on the biology and fitness of fall
369 armyworm, *Spodoptera frugiperda*, a recently invaded pest of maize in India. *Silicon*.
370 <https://doi.org/10.1007/s12633-020-00901-8>

371 Nascimento, A.M., Assis, F.A., Moraes, J.C., & Sakomura, R. (2014). Feeding non preference
372 of *Spodoptera frugiperda* (Lepidoptera: Noctuidae) induced by silicon application in rice crop.
373 *Agraria* 9(2), 215-218. <https://doi.org/10.5039/agraria.v9i2a3930>

374 Nascimento, A.M., Assis, F.A., Moraes, J.C., & Souza, B.H.S. (2017). Silicon application
375 promotes rice growth and negatively affects development of *Spodoptera frugiperda* (J.E.
376 Smith). *Journal of Applied Entomology* 142, 241–249. <https://doi.org/10.1111/jen.12461>

377 Rahman, A., Wallis, C.M., & Uddin, W. (2015). Silicon-Induced Systemic Defense Responses
378 in Perennial Ryegrass Against Infection by *Magnaporthe oryzae*. *Phytopathology* 105, 748–
379 757. <https://doi.org/10.1094/PHYTO-12-14-0378-R>

380 Rémus-Borel, W., Menzies, J.G., & Bélanger, R.R. (2005). Silicon induces antifungal
381 compounds in powdery mildew-infected wheat. *Physiological and Molecular Plant Pathology*
382 66, 108–115. <https://doi.org/10.1016/j.pmpp.2005.05.006>

383 Reynolds, O.L., Keeping, M.G., & Meyer, J.H. (2009). Silicon-augmented resistance of plants
384 to herbivorous insects: A review. *Annals of Applied Biology* 155, 171–186.
385 <https://doi.org/10.1111/j.1744-7348.2009.00348.x>

386 Reynolds, O.L., Padula, M.P., Zeng, R., & Gurr, G.M. (2016). Silicon: potential to promote
387 direct and indirect effects on plant defense against arthropod pests in agriculture. *Frontiers in*
388 *Plant Science* 7,744. <https://doi.org/10.3389/fpls.2016.00744>

389 Richmond, K.E., & Sussman, M. (2003). Got silicon? The non-essential beneficial plant
390 nutrient. *Current Opinion in Plant Biology* 6, 268–272. [16](https://doi.org/10.1016/S1369-</p></div><div data-bbox=)

391 5266(03)00041-4

392 Rostás, M. (2007). The effects of 2,4 dihydroxy-7-methoxy-1,4-benzoxazin-3-one on two
393 species of *Spodoptera* and the growth of *Setosphaeria turcica* in vitro. *Journal of Pest Science*
394 80, 35-41. <https://doi.org/10.1007/s10340-006-0151-8>

395 Silva, A.A., Alvarenga, R., Moraes, J.C., & Alcantra, E. (2014). Biology of *Spodoptera*
396 *frugiperda* (J.E. Smith) (Lepidoptera: Noctuidae) in colored cotton treated with silicon.
397 *EntomoBrasilis* 7(1), 65–68. <https://doi.org/10.12741/ebrasilis.v7i1.365>

398 Strömberg, C.A.E., Di Stilio, V.S., & Song, Z. (2016). Functions of phytoliths in vascular
399 plants: an evolutionary perspective. *Functional Ecology* 30, 1286–1297.
400 <https://doi.org/10.1111/1365-2435.12692>

401 Wang, Z., Zhu, W., Chen, F., Yue, L., Ding, Y., Xu, H., Rasmann, S., & Xiao, Z. (2021).
402 Nanosilicon enhances maize resistance against oriental armyworm (*Mythimna separata*) by
403 activating the biosynthesis of chemical defenses. *Science of the Total Environment* 778,
404 146378. <https://doi.org/10.1016/j.scitotenv.2021.146378>

405 Waterman, J.M., Hall, C.R., Mikhael, M., Cazzonelli, C.I., Hartley, S.E., & Johnson, S.N.
406 (2021). Short-term resistance that persists: Rapidly induced silicon anti-herbivore defence
407 affects carbon-based plant defences. *Functional Ecology* 35(1), 82-92.
408 <https://doi.org/10.1111/1365-2435.13702>

409 Yang, L., Han, Y., Li, P., Li, F., Ali, S., & Hou, M. (2017). Silicon amendment is involved in
410 the induction of plant defense responses to a phloem feeder. *Scientific Reports* 7, 4232.
411 <https://doi.org/10.1038/s41598-017-04571-2>

412 Zheng, X.-L., Cong, X.-P., Wang, X.-P., & Lei, C.-L. (2011). A review of geographic
413 distribution, overwintering and migration in *Spodoptera exigua* Hübner (Lepidoptera:
414 Noctuidae). *Journal of the Entomological Research Society* 13(3), 39-48.

415 Figures

416

417

418

419

