

Molecular effective quantification of pathogens and total flora in meat products

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Objectives

The aim of our study was to develop an effective molecular quantification for different flora of interest for food industry.

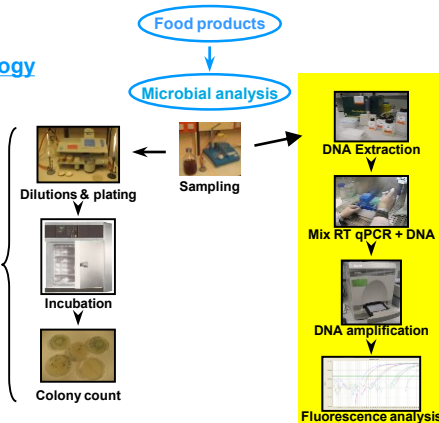
Introduction

Classical analysis for food shelf-life uses traditional microbiological techniques. Those are long, heavy and tedious. There is a demand from our industrial partners to develop a faster and easier alternative technique. Real time quantitative PCR (RT qPCR) has recently entered service in the field of food science and technology. Several methods have been developed for the detection and quantification of opportunistic pathogens and certain non-pathogenic spoilage microorganisms in food products. All molecular techniques needed an enrichment step which could hedge the quantification.

Classical microbiology

≈ 30 h
for ≈ 30 samples

Tedious
Heavy
cheap



RT qPCR

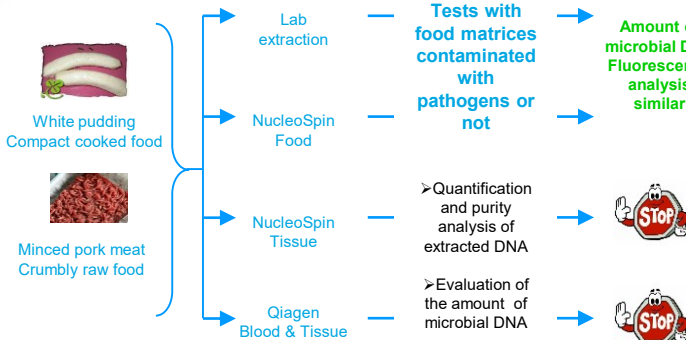
≈ 7 h
for 2 x 96 samples

Expensive
Fast
DNA samples
could be stored
Analysis could
be delayed

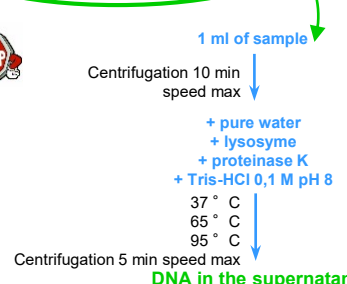
Challenges & Results

1 Extraction

➤ Efficient for all food matrices

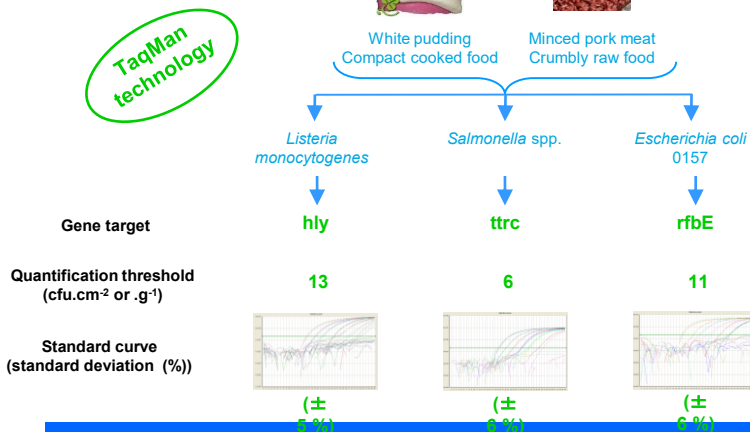


Faster
Cheaper
Effective on different food matrices



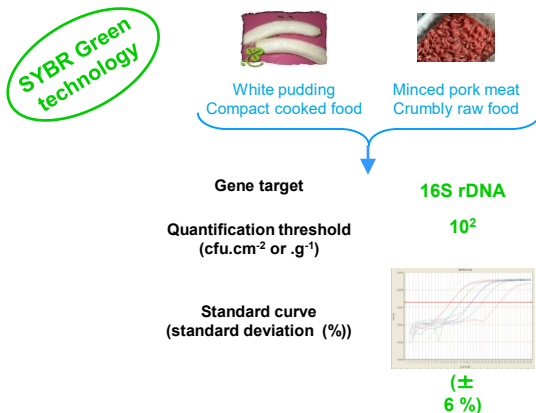
2 For pathogens

➤ Enrichment needed
➤ Possibility of quantification decreased or even impossible



3 Total flora

➤ Composition varied according to tested food
➤ Original flora very diversified



Conclusions

For our industrial partners products, a microbial DNA extraction has been developed. This extraction could be applied on cooked and raw pork meat. Effective food pathogens and total flora quantification protocols has been performed. Standard curves have a low standard deviations, around ± 6 %. Quantification threshold for pathogens is around 10 cfu.cm⁻² or .g⁻¹, and below, a detection is possible. The protocol of RT qPCR is a very interesting option for food industry, but more food matrices should be tested.