ENHANCEMENT OF CONCOMITANT IMMUNITY AFTER RADIATION THERAPY AND IMMUNOTHERAPY IN A SYNGENEIC MURINE TUMOUR SYSTEM

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The efficacy of radiation therapy of tumours depends on lethal effects on malignant cells but also on modifications of vessels and connective tissue (STENSTRÖM et coll. 1955, PUCK & MARCUS 1956, HEWITT & WILSON 1959). Tumour irradiation induces an immune response of the host cooperating in the destruction of allogeneic malignant cells (DENEUFBOURG 1972, 1975). These preliminary findings were tested in an experiment system consisting of a chemically induced epidermoid carcinoma transplanted in a syngeneic situation. The influence of irradiation and active non-specific immunotherapy was assessed in vivo on the level of concomitant immunity. The results are now reported.

Materials and Methods

Inbred female CBA mice 2 to 3 months old were used. They were fed with standard pellet diet and water ad libitum. The experiment groups contained 30 to 40 mice for each data point.

An epidermoid carcinoma was induced by repeated painting of the skin with 20-methylcholanthrene, and a syngeneic tumour was maintained by serial subcutaneous transplantations. Tumour pieces aseptically prepared were inoculated intradermally in the median dorsal region of the animals. The tumour growth followed a linear progression and remained purely local without deep infiltration or visible metastases. Host mean survival time was about 30 days.

Irradiation was performed on day 8 of tumour growth. An original set-up of holding and shielding allowed a good exposure of the target volume without anesthesia (DENEUFBOURG 1972). The irradiation was delivered through two parallel opposed portals and conferred single doses of 25 or 5 Gy at mid-depth. The radiation was generated with a Philips apparatus at 50 kV and 2 mA with a 1 mm Al filter and a focus-skin distance of 4 cm. The whole-body dose did not exceed one per cent of the tumour dose.

Active non-specific immunotherapy. On day 8 of tumour growth 0.5 mg of a suspension of Corynebacterium parvum (2 mg/ml; kindly supplied by Institut Méreux, Lyon, France) was injected intraperitoneally or subcutaneously in each animal.

Assessment of concomitant immunity. Three groups of mice (non-irradiated, irradiated, or after immunotherapy) bearing a dorsal tumour received a second intradermal tumour implant on their ventral side. This challenge graft contained about $10^6$ tumour cells; it was observed 3 times weekly and rejections were recorded.

Results

Tumour growth. After a latent period of at least 3 days anti-tumour immunization rose to 31 per cent on day 6 and to 73 per cent on day 8. Then it
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Fig. 1. Rejection rate of a challenge graft (in per cent) implanted on days 3, 6, 8, 10 and 15 of primary tumour growth and influence of radiation therapy (25 and 5 Gy tumour dose on day 8). No treatment (□), 25 Gy (□□), 5 Gy (□□□).

Table

Concomitant immunity during the growth of a syngeneic epidermoid carcinoma and influence of tumour irradiation and immunotherapy. Rejection rate of a challenge graft implanted on days 3, 6, 8, 10 and 15 of primary tumour progression. Treatment on day 8: 25 Gy and 5 Gy tumour dose, intraperitoneal or subcutaneous injection of Corynebacterium parvum.

<table>
<thead>
<tr>
<th>Rejection rate of challenge graft</th>
<th>Day 3</th>
<th>Day 6</th>
<th>Day 8</th>
<th>Day 10</th>
<th>Day 15</th>
</tr>
</thead>
<tbody>
<tr>
<td>No treatment</td>
<td>0%</td>
<td>31%</td>
<td>73%</td>
<td>55%</td>
<td>43%</td>
</tr>
<tr>
<td></td>
<td>(0/34)</td>
<td>(11/35)</td>
<td>(22/30)</td>
<td>(21/38)(^a)</td>
<td>(15/35)(^b)</td>
</tr>
<tr>
<td>25 Gy</td>
<td>–</td>
<td>–</td>
<td>73%</td>
<td>82%</td>
<td>95%</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>(22/30)</td>
<td>(31/38)(^a)(^c)</td>
<td>(38/40)(^b)(^d)</td>
</tr>
<tr>
<td>5 Gy</td>
<td>–</td>
<td>–</td>
<td>73%</td>
<td>45%</td>
<td>36%</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>(22/30)</td>
<td>(17/38)(^a)</td>
<td>(14/39)(^d)</td>
</tr>
<tr>
<td>Corynebacterium parvum, intraperitoneally</td>
<td>–</td>
<td>–</td>
<td>73%</td>
<td>65%</td>
<td>75%</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>(22/30)</td>
<td>(26/40)(^f)</td>
<td>(24/32)(^e)</td>
</tr>
<tr>
<td>Corynebacterium parvum, subcutaneously</td>
<td>–</td>
<td>–</td>
<td>73%</td>
<td>54%</td>
<td>53%</td>
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<tr>
<td></td>
<td></td>
<td></td>
<td>(22/30)</td>
<td>(20/37)</td>
<td>(16/30)(^f)</td>
</tr>
<tr>
<td>5 Gy and Corynebacterium parvum, intraperitoneally</td>
<td>–</td>
<td>–</td>
<td>73%</td>
<td>88%</td>
<td>72%</td>
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<td></td>
<td></td>
<td></td>
<td>(22/30)</td>
<td>(13/35)(^f)(^g)</td>
<td>(26/36)(^h)</td>
</tr>
<tr>
<td>5 Gy and Corynebacterium parvum, subcutaneously</td>
<td>–</td>
<td>–</td>
<td>73%</td>
<td>59%</td>
<td>44%</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>(22/30)</td>
<td>(22/37)(^k)</td>
<td>(15/34)(^h)</td>
</tr>
</tbody>
</table>

Statistical analysis of results:
\(^a\) p<0.01. \(^b\) p<0.001. \(^c\) p<0.01. \(^d\) p<0.001. \(^e\) p<0.05. \(^f\) p<0.02. \(^g\) p<0.01. \(^h\) p<0.02.
gradually declined and the rejection rate fell to 55 per cent on day 10 and 43 per cent on day 15 (Table, Figs 1, 2).

Irradiation. The level of concomitant immunity increased after 25 Gy to 82 per cent 2 days later and to 95 per cent 7 days later, after 5 Gy the rejection rate of the challenge graft amounted to 45 per cent and 36 per cent, respectively.

Immunotherapy. After intraperitoneal administration of Corynebacterium parvum on day 8 of tumour growth concomitant immunity reached 65 per cent on day 10 and 75 per cent on day 15. Using the subcutaneous route the corresponding rejection rates were 54 per cent and 53 per cent, respectively.

Immunotherapy and irradiation. A 5 Gy tumour dose associated with intraperitoneal injection of Corynebacterium parvum induced an 88 per cent rejection rate of the challenge graft on day 10. Antitumour resistance evaluated on day 15 was present in 72 per cent of the mice. Results with adjuvant immunotherapy via the subcutaneous route were similar to the controls: 59 and 44 per cent, respectively.

Discussion

The efficacy of radiation therapy depends to a large extent on the lethal effects on tumour cells (PUCK & MARCUS, HEWITT & WILSON). Alterations of connective tissue and vascular modifications of the tumour bed are also known to play a part (STENSTROM et coll.).

Previously it was suggested that tumour irradiation induces an immune response of the host cooperating in the destruction of allogeneic malignant cells (DENEUFBOURG 1972, 1975). These preliminary results were confirmed in the present experiments using a chemically induced epidermoid carcinoma in a mouse syngeneic system.

An antigenic tumour while progressively growing induces an immunization of the host cooperating in the destruction of allogeneic malignant cells (DENEUFBOURG 1972, 1975). These preliminary results were confirmed in the present experiments using a chemically induced epidermoid carcinoma in a mouse syngeneic system.

According to the present results, tumour irradiation reverses the vanishing phase of concomitant immunity. A 25 Gy single dose was effective within 48 hours and conferred anti-tumour resistance to 95 per cent of the animals 8 days later. This reaction was dose related.

Data are still scarce in the literature about such immunologic effects. Fractionated tumour irradiation with a total dose of 90 to 120 Gy induced recovery of concomitant immunity in mice bearing a transplantable mammary tumour (BARSKI et coll. 1974). Reappearance of immune resistance against a fibrosarcoma was observed after a tumour dose of 60 Gy but was delayed as long as vascular connections between host and tumour persisted (VAAGE 1973). In both reports it was noted that a comparable level of immunity was achieved after surgery and that an incomplete tumour resection prevented full recovery. In the present system, irradiation did not lead to tumour elimination and yet reversed the decline of concomitant immunity without delay.

Mechanisms of radiation induced immune response are presently under investigation. Cell membrane destruction by irradiation is known to release antigenic material (MOROSON 1978). Irradiation may induce antigenic modifications or unmasking and enhance host immunization (MARUYAMA 1968). Untrapping of cytotoxic antibodies might also occur. Destruction of suppressor cells by whole-body exposure as a side-effect of tumour treatment is unlikely, due to the irradiation procedure.

Adjuvant active non-specific immunotherapy with Corynebacterium parvum counteracted the decline of concomitant immunity associated with tumour progression. Immunostimulation was effective within 48 hours and lasted 8 days at least. The discrepancy of activity between intraperitoneal and subcutaneous injections (CASTRO 1977) was confirmed. Irradiation at low dosage and Corynebacterium parvum acted synergistically. An irradiation with 5 Gy is unable to modify the tumour host balance but potentiates non-specific immunotherapy. Antigen availability may be a prerequisite for full range response to immunostimulation.

Detrimental immunosuppressive effects of radiation therapy have been pointed out especially in breast carcinoma treatment (STJERNwäRD 1974). However, the relations are perhaps more complicated than are presumed. The present results emphasize the fact that appropriate tumour irradiation and immunostimulation may also exert a beneficial influence on host resistance.
SUMMARY

Concomitant immunity was evaluated in vivo towards a chemically induced epidermoid carcinoma transplanted in a syngeneic situation. Radiation therapy reversed the declining phase of concomitant immunity associated with tumour progression. The rejection rate of a challenge graft amounted to 82 and 95 per cent, respectively, 2 and 8 days after 25 Gy as compared with 55 and 43 per cent in unirradiated controls. Radiation induced immune recovery was dose related and proved to be different from restoration of immunity following surgical removal of the tumour. Immunotherapy with intraperitoneal injection of Corynebacterium parvum significantly improved concomitant immunity. Immunostimulation and irradiation at low dosage act synergistically on host anti-tumour resistance.

ACKNOWLEDGEMENTS

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REFERENCES

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