ESR SPECTROSCOPY FOR THE STUDY OF AN INFLAMMATION-INDUCED **AKI** CELLULAR MODEL



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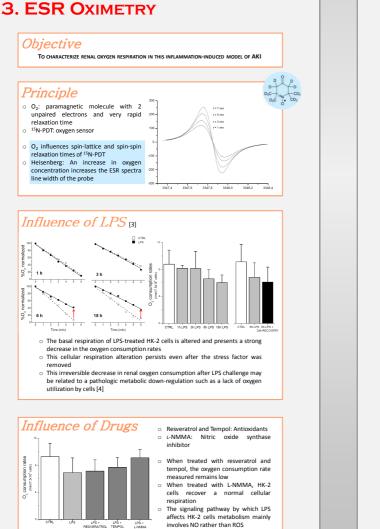


1. BACKGROUND

Sepsis can be considered as a heterogeneous disease process generated by a complex interaction of pathogen and host inflammatory response. It may lead to organ dysfunction distant from the primary site of infection and cause downstream effects such as multiple organ failure [1].

The kidney is one of the target organs of sepsis which is well-known to be a risk factor for the development of acute kidney injury (AKI). Recent research activities in the mechanisms involved in the development of AKI in sepsis emphasize the central role of hemodynamic and inflammatory events.

More particularly, two mechanisms are suggested to explain the inability of the injured kidney to extract oxygen: tissue hypoxia and cellular energetic metabolism dysfunction [2]. Our working hypothesis of the pathophysiology of AKI is based on cellular respiratory dysfunction due to the inflammatory response inherent to sepsis.



2. IN VITRO MODEL

We developed an *in vitro* model of inflammation-induced acute kidney injury using HK-2 cells exposed to lipopolysaccharide (LPS) [3].

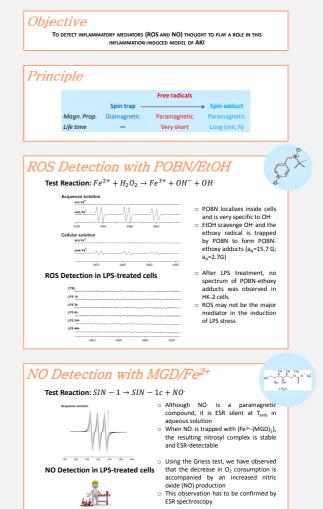


HK-2 cells, derived from human proximal tubular cell (PTC) of a human kidney, were cultured in DMEM supplemented with 10% FBS, 2 mM L-Glutamine, 100 U/ml Penicillin and 100 μ g/ml Streptomycin.

To stimulate the pro-inflammatory sate of PTC, cells were exposed to LPS from *E. coli* 055:85. LPS is released from the gram-negative bacteria and is one of the major initiators inflammatory response during sepsis. LPS is known to produce an early rise in cytokines through activation of Tolllike receptor.

One concentration (1 $\mu g/ml)$ and different incubation time (1h to 24h) were tested.

4. SPIN TRAPPING



CONCLUSIONS

Overall, ESR spectroscopy and the model of HK-2 cells exposed to LPS display some key features of inflammationinduced acute kidney injury:

- 1. Alteration in the renal cellular respiratory function
- 2. Role of NO· rather than ROS in the signalling pathway
- 3. Role of INOS in the generation of NO-

OPEN QUESTIONS

- Since L-NMMA permitted the HK-2 cells to recover a normal cellular respiration, does it blocked NO generation?
- Is it possible to detect the activation of inducible NO synthase?
- Are mitochondrial alterations the mechanism of HK-2 cells basal respiration perturbations during inflammation?

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REFERENCES

 R.W. Schrier, W. Wang, New England Journal of Medicine 351 (2004) 159-169.
A. Haros, O. Huet, J. Duanteus, Current Opinion in Anesthesiology 22 (2009) 143-149.
C. Qualin, A. Moulthy-Mickalad, Duanteus, G. Saller, M. Noebeke, Biochemical and Biophysical research Communications 423 (2012) 350-354.
R.J. Levy, Shock 22 (2007) 24-24.