Specificity of different RIA systems for measurement of bovine pregnancy-associated glycoproteins against carbohydrates and placental hormones

A. Ayad¹, N.M. Sousa¹, J. Sulon¹, E. Clerget¹, M. Iguer-Ouada² and J. F. Beckers¹

¹ Department of Physiology of Animal Reproduction, Faculty of Veterinary Medicine, University of Liege, B-4000 Liege, Belgium ² Department of Organisms and Populations Biology, Faculty of Nature and Life Sciences, University A. Mira, 06000, Bejaia, Algeria

Antroduction

The pregnancy-associated glycoproteins (PAG) constitute a large family of glycoproteins specifically expressed in the outer epithelial cell layer of the placenta in eutherian species. They are members of the aspartic proteinase family (Green et al., 2000).

Radioimmunoassay for PAG detection in serum or plasma samples is currently used as a specific serological method for pregnancy diagnosis in cattle (Zoli et al., 1992).

Different numbers of N-glycosylation sites of asparagines have been observed in PAG sequences deduced from cDNA or obtained after N-terminal microsequencing (Xie et al., 1997a). Note that the placental glycoproteins from human (hCG) and equine origins (PMSG) have also been shown to contain several lateral sugar chains.

> The aim of this study was to test the specificity of five PAG-RIA systems against different carbohydrates and placental hormones.

Materials and methods

- > The following carbohydrate preparations of commercial origin were used to test the specificity of each RIA: N-acetyl-D-galactosamine, N-acetyl-D-glucosamine, N-acetyl-neuraminic acid, D(+)-lactose, D(+)-glucose, Methyl- α -D-mannopyranoside, Mannitol, and D(+)-galactose. Furthermore, the specificity was tested against the following hormones: PMSG and hCG.
- ➤ Each carbodrate was dissolved and diluted in Tris- BSA buffer to obtain the following concentrations: 1.0, 10², 10⁴ and 10⁶ ng/ml. For PMSG and hCG, the tested concentrations were 10⁻³, 10⁻¹, 10¹ and 10³ IU/ml.
- ➤ Polyclonal antisera were collected from rabbits immunized (R#) against different PAG preparation according to the technique of Vaitukaitis et al. (1971): R#497 was raised against boPAG67, R706 against caPAG55+62, R#780 against ocPAG57+59, R#809 against ovPAG55.
- > These four antisera were mixed (R#497 one part; R#706 one part; R#780 two parts; and R#809 two parts) and used as an additional antiserum (Pool) (Ayad et al., 2006).
- > The measurement were performed according to a modified method of Perényi et al. (2002b).
- > A dilution of a product was considered as crossing-reacting if the determined B/Bo value lower than the MDL/Bo value.

In RIA-809, there were weak inhibition B/B0 (%mean \pm SD) by N-acetylneutraminic acid and hCG when tested in the concentration of 1 mg/mL (88.11 \pm 5.34) and 1000 UI/mL (93.89 \pm 0.84), respectively.

Faculté de médecine

The other four RIA systems did not give any cross-

Carbohydrates tested	B/B ₀ binding interference at the following concentrations (ng/mL)																			
	RIA-497				RIA-706					RIA	-780		RIA-809					RIA-Pool		
	1	10²	10 ⁴	10 ⁶	1	10²	104	10 ^ε	1	10²	104	10 ^ε	1	10 ²	10 ⁴	10 ⁶	1	10²	10 ⁴	10
N-acetyl-D-galatosamine	-	-	-	-		-	-	-	-	-	-	-		-	-	-		-	-	-
N-acetyl-D-glucosamine	-	-	-	-		-	-	-		-	-	-		-	-	-		-	-	-
N-acetylneuraminic acid		-	-	-		-	-	-	-	-	-	-		-	- (-	-	-
D(+)-Lactose		-	-				-					-			-			-	-	-
D(+)-Glucose		-	-	-		-	-	-	-	-	-	-		-	-	-		-	-	-
Methyl-α-D-mannopyranoside		-	-	-		-	-	-	-	-	-	-		-	-	-		-	-	-
Mannitol		-	-	-	-	-		-	-	-	-	-		-	-	-		-	-	-
D(+)-Galactose																				

Placental protein tested	_	RIA	-497		В/1		-706	ence	at the		-780	conc	entra	RIA	•	nL)		RIA-	Pool	
	10-3			10 ³	10-3	10-1	101	10 ³	10-3			10 ³	10-3			10 ³	10-3		101	10 ³
PMSG	-	-	-	-		-	-	-	-	-		-	-	-		-		-		-
hCG	-			-		-	-	-	٠			-		-	- 1	①) -	-		-
																				_

Conclusion

In conclusion, four RIA systems can be considered as specific for the detection of PAG concentrations in palsma or serum.

Only N-acetylneutraminic acid and hCG could decrease the binding of the radiolabelled PAG tracer to the antibodies, and were able to slightly cross-react with the antiserum 809 used.

References

- Nyad A, Sousa NM, Sulon J, Iguer-Ouada M and Beckers JF. Comparison of five radioimmunoassay systems for PAG measurement: ability to detect early pregnancy in cow. Reprod. Dom. Anim. 2006 (submitted).
- Green JA, Xie S, Quan X, Bao B, Gan X, Mathialagan N, Beckers JF, Roberts RM. Pregnancy-associated bovine and ovine glycoproteins exhibit spatially and temporally distinct expression patterns during pregnancy. Biol. Reprod. 2000; 62: 1624-1631.
 Perényi Z, Szenci O, Sulon J, Drion PV, Beckers JF. Comparison of the
- Perényi Z, Szenci O, Sulon J, Drion PV, Beckers JF. Comparison of the ability of three radioimmunoassays to detect pregnancy-associated glycoproteins in bovine plasma. Reprod. Dom. Anim. 2002b; 37: 100-104.
- Vaitukaitis J, Robbins JB, Nieschlag E, Ross GT. A method for producing specific antisera with small doses of immunogen. J. Clin. Endorinol. Metab. 1971; 33: 988-991.
- Xie S, Green J, Bao B, Beckers JF, Valdez KE, Hakami L, Roberts RM. Multiple pregnancy-associated glycoproteins are secreted by day 100 ovine placental tissue. Biol. Reprod. 1997a; 57: 1384-1393.
- Zoli AP, Guilbaut LA, Delahaut P, Benitez Ortiz W, Beckers JF. Radioimmunoassay of a bovine pregnancy-associated glycoprotein in serum: its application for pregnancy diagnosis. Biol. Reprod. 1992; 46: 83-92.